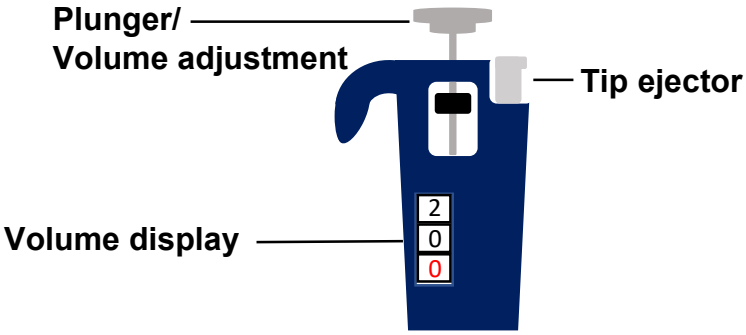
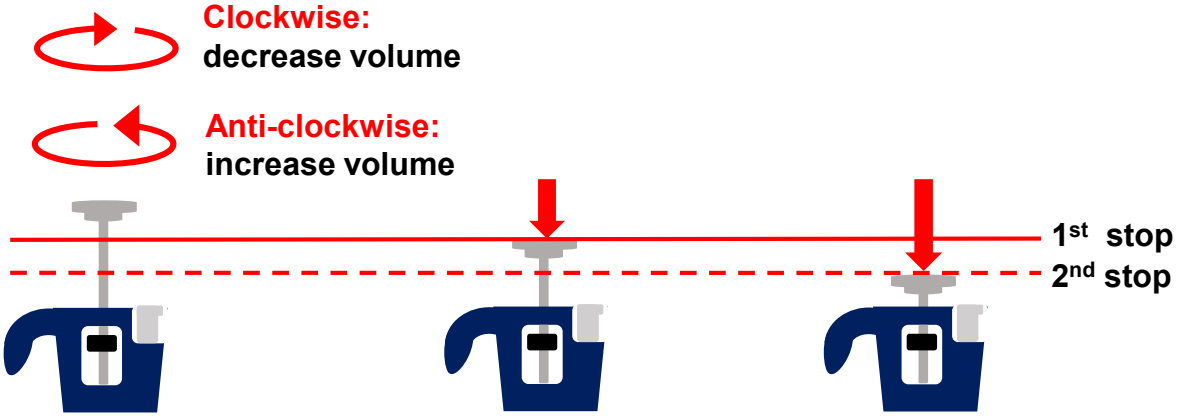
How to use a micropipette?





To adjust volume

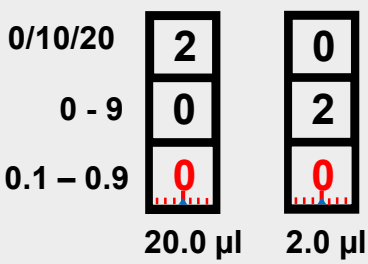
To release most of the liquid

To release last little bit

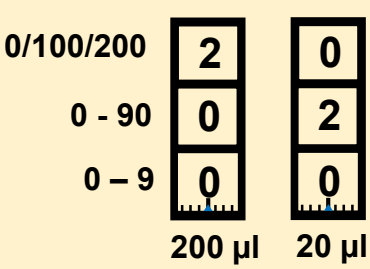
Do NOT over turn the plunger!

① Set Volume

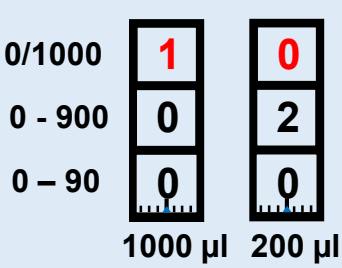
P 20 pipette = 2 – 20 µl



P 200 pipette = 20 – 200 µl



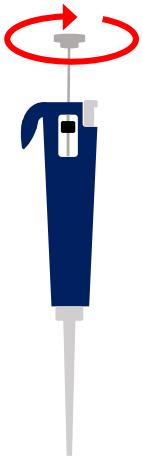
P 1000 pipette = 200 – 1000 µl



Never set volume above or below the limits!

② Step of pipetting liquid

a. Set volume



b. Insert pipette tip

Tip holder

Never touch its tip with anything!

Push hard on the Pipette tip to make it tight

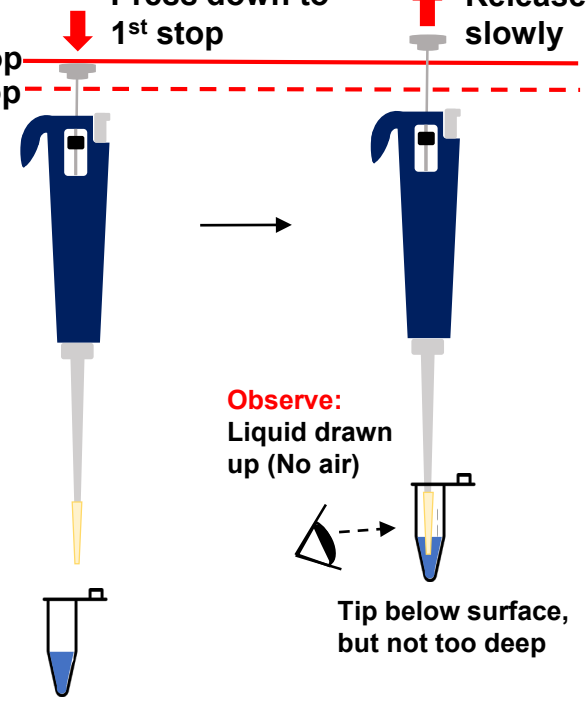
c. Suck up liquid

Press down to 1st stop

Release slowly

1st stop

2nd stop



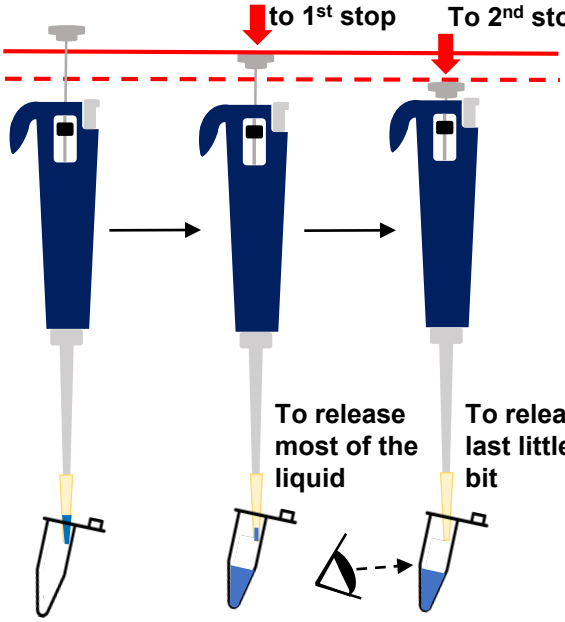
Observe:
Liquid drawn up (No air)

Tip below surface, but not too deep

d. Release liquid

Press down to 1st stop

***Press down To 2nd stop**



To release most of the liquid

To release last little bit

***When adding liquid in wells of the gel, press down to 1st stop only. Pressing to 2nd stop will eject air, making the liquid in the well spill out.**

e. Lift up the tip

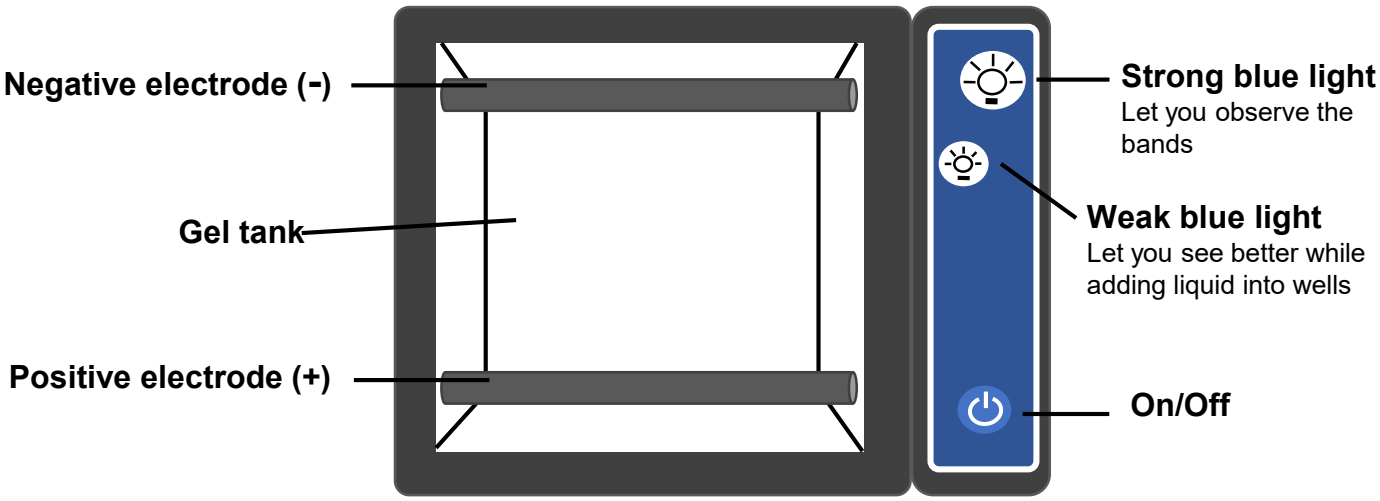
Keep pressing at 2nd stop while lifting up

f. Eject the tip

Release the plunger slowly and press tip ejector

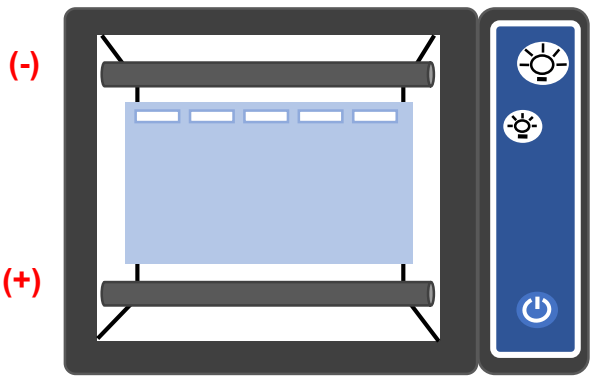
Replace a new tip for another liquid

How to do a gel electrophoresis?



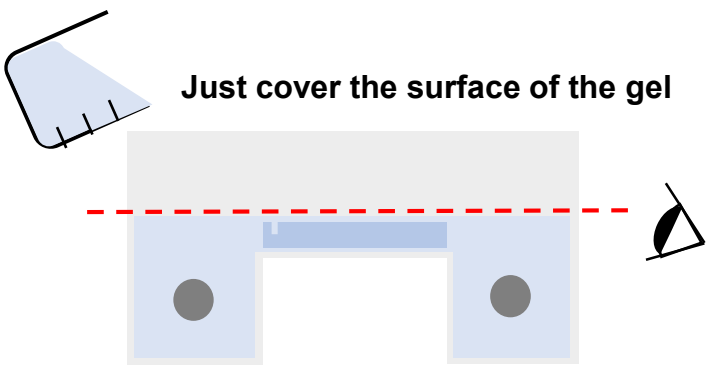
Gel System

a. Put the gel into the gel tank



Make sure the wells are on the negative (-) side

b. Add buffer to the gel tank

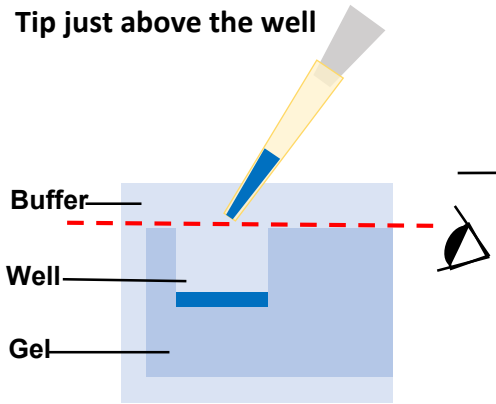


How to do a gel electrophoresis?

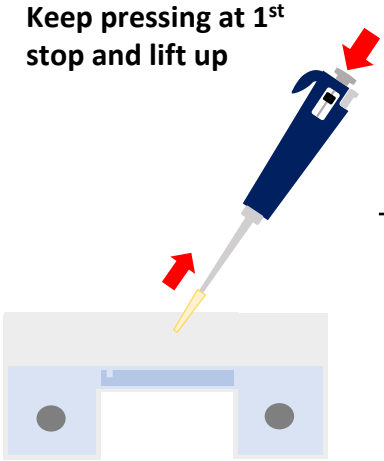
c. Add solution into the wells

Press to 1st Stop only

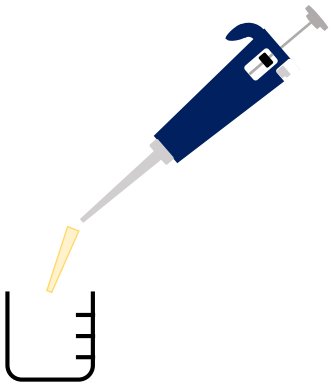
Tip just above the well



Keep pressing at 1st stop and lift up

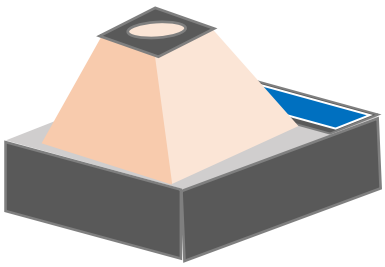


Add another solution with a new tip



Press weak blue light to make the wells easier to see

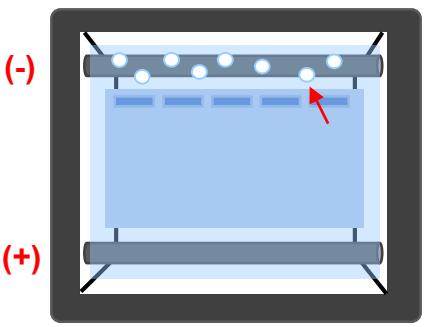
d. Turn on power



Place the photo hood on the gel system

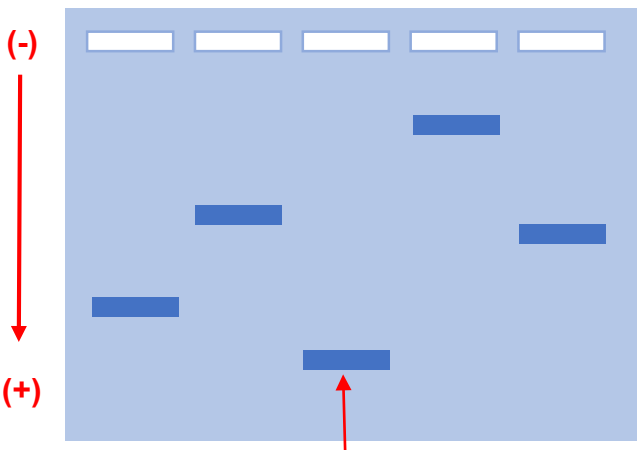
Press strong blue light to observe the bands

e. Start running the gel



Bubbles come out at negative (-) electrode

f. Observe bands appearing from (-) to (+) electrode



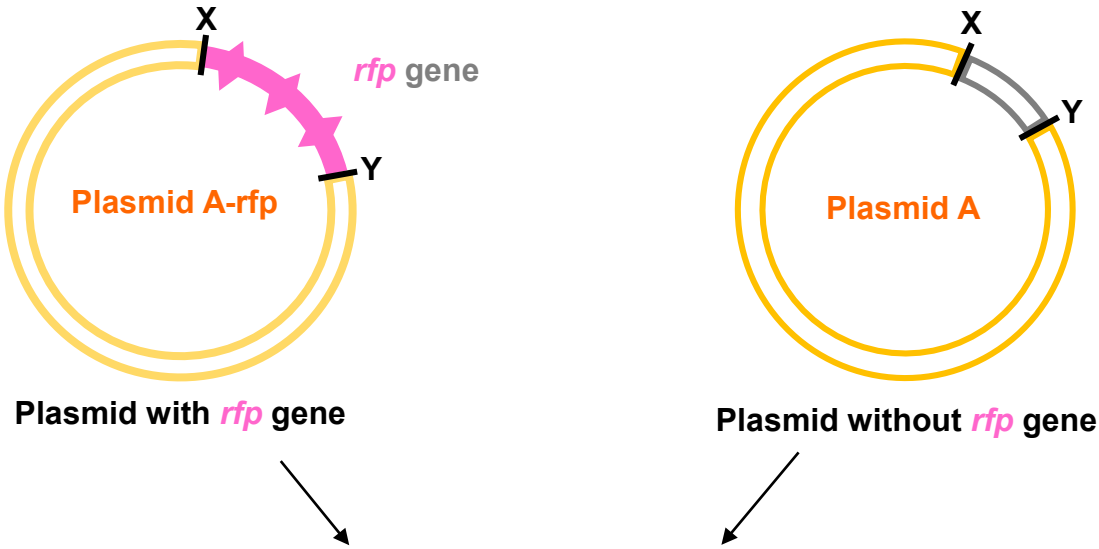
Larger (move slower)

Fragment Size

Smaller (move faster)

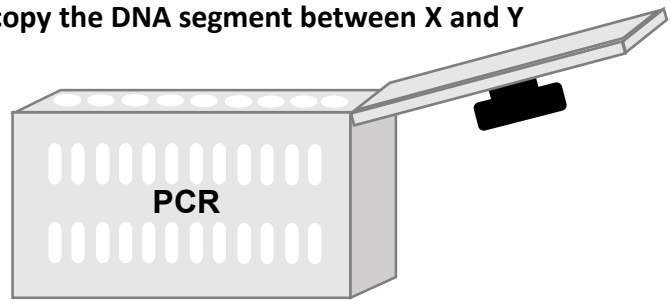
Bands show different sizes of DNA fragments

How to check if a plasmid has the *rfp* gene?



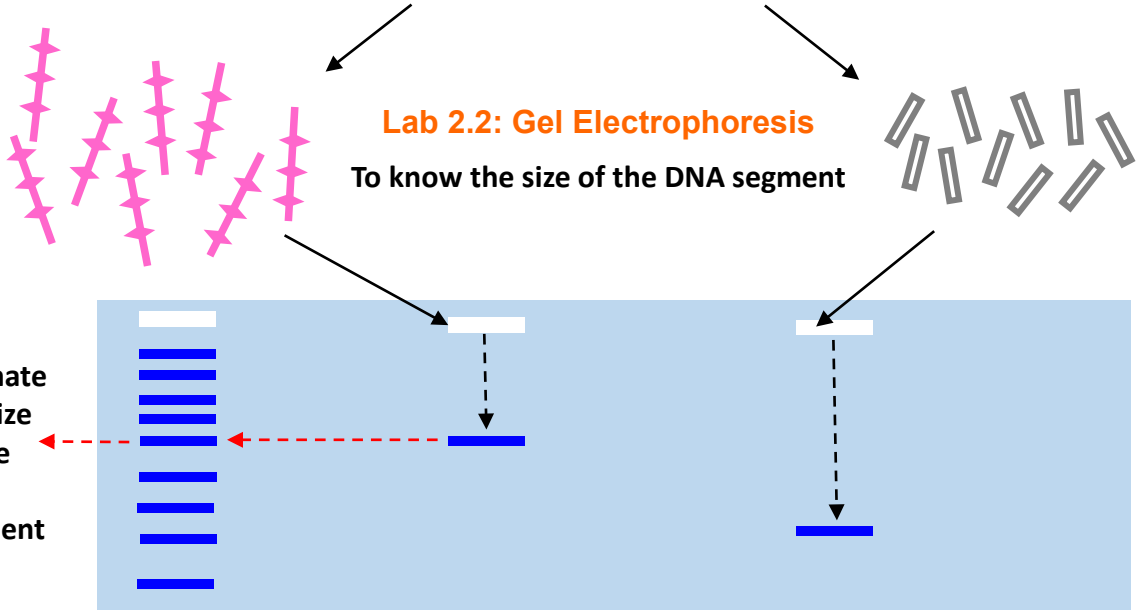
Lab 2.1: Polymerase Chain Reaction (PCR)

To copy the DNA segment between X and Y



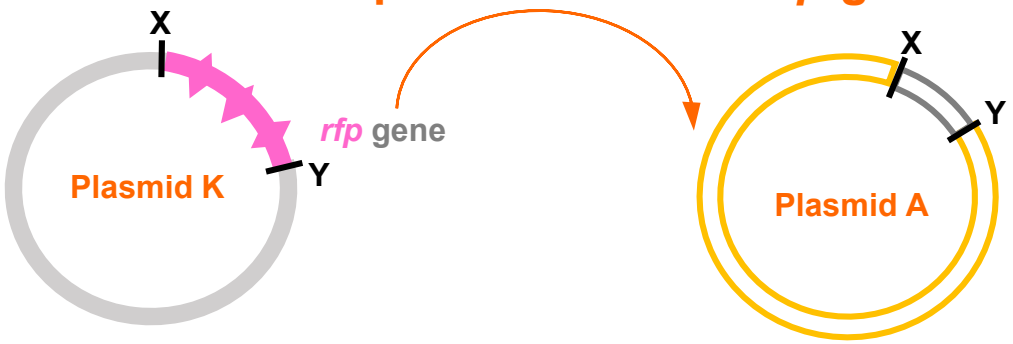
Lab 2.2: Gel Electrophoresis

To know the size of the DNA segment



If the size is similar to the *rfp* gene, it is concluded that the *rfp* gene is in plasmid A

How to make a plasmid with the *rfp* gene?

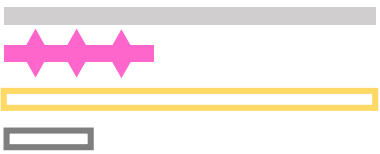


Source of *rfp* gene

Plasmid with DNA to express *rfp* gene and make bacteria resistant to ampicillin. So we know which bacteria have got the plasmid.
+ Restriction Enzymes

Lab 3.1: Restriction Digestion

To cut the plasmid at site X and Y

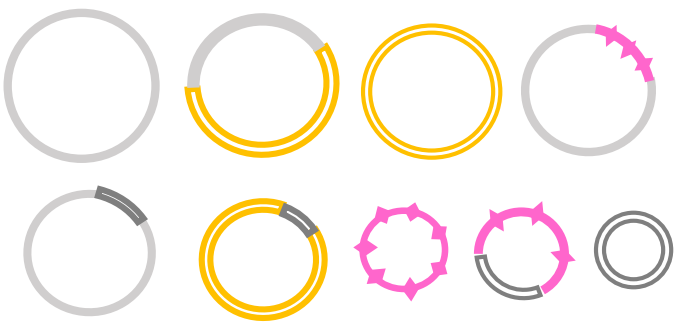


Lab 3.2: Ligation

To stick the *rfp* gene to plasmid A between site X and Y

Mix the DNA fragments
+ DNA Ligase

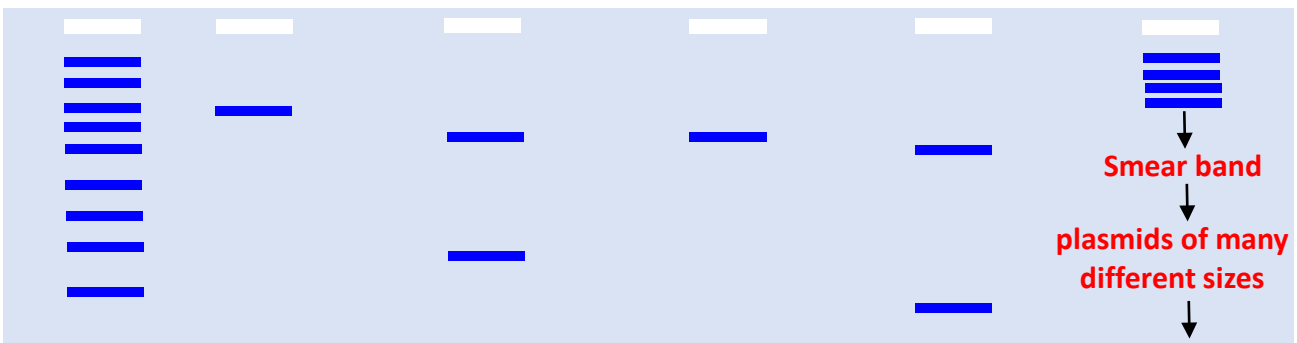
Target: Plasmid A with *rfp* gene



Other useless ligated plasmids

Lab 3.3: Gel Electrophoresis

To check if ligation occurs

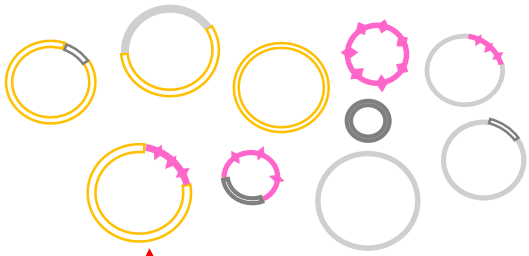


DNA Ladder | Uncut Plasmid K | Cut Plasmid K | Uncut Plasmid A | Cut Plasmid A | Ligation happens

How to put the plasmid with *rfp* gene into a bacteria?

Plasmid A with *rfp* gene

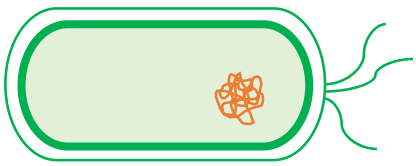
O
R



Ligated plasmids
(include plasmid A with *rfp* gene)

Lab 4: Transformation

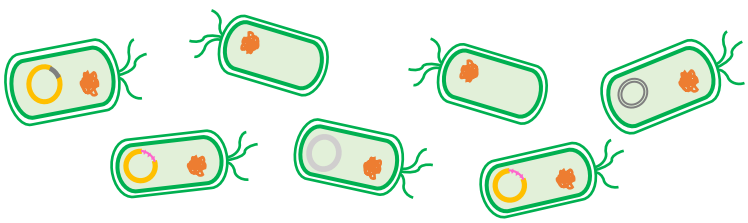
To put the plasmid A with *rfp* gene into bacteria



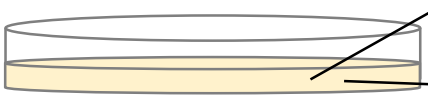
E. coli bacteria

Heat Shock

To open pores on the cell membrane so that plasmids can be taken into the bacteria.



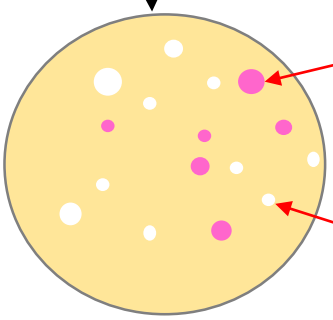
E. coli bacteria
(some take in plasmids, some do not)



Bacterial culture
grow individual bacteria into colonies

Ampicillin
kill any bacteria without plasmid A that gives the bacteria resistance to ampicillin

Arabinose
Express the *rfp* gene and produce red fluorescent protein



Red bacterial colony –
bacteria that have taken in plasmid A with *rfp gene*

White bacterial colony –
bacteria that have taken in plasmids with ampicillin resistance but no *rfp gene*